



Evaluation of acid permanganate extraction as an index of soil nitrogen availability

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Summary A chemical measure of soil nitrogen (N) availability has been evaluated. The method involved the estimation of initial mineral N, plus mineralizable N released with 0.05 N KMnO_4 in $1\text{ N H}_2\text{SO}_4$. The results obtained correlated highly significantly with the N uptake by wheat plants ($r = 0.72, P < 0.01$).

Introduction

The assessment of potentially available soil N by the biological methods involving soil incubation under controlled laboratory conditions have been reported to be the most satisfactory methods¹. Although these methods are quite reliable they are time consuming. Therefore, research efforts have been made to find out a rapid chemical method that could provide reliable index of soil N availability. A review by Dahnke and Vasey⁵ mentioned 17 chemical methods that have been used for testing soils for available N. According to Fox and Piekielek⁶, out of these 17 methods, only initial $\text{NO}_3\text{-N}$ and soil organic matter are used by a few soil testing laboratories to predict the N supplying capacity of soils. Some of the recent studies indicated that $\text{NH}_4\text{-N}$ released from soil organic matter with alkaline or acid permanganate could be used as an index of soil nitrogen availability^{9,10,12}. Work carried out in this laboratory⁷ has also indicated acid permanganate extraction to be a simple and rapid measure of available N in soils. This index has been proposed on the basis of results for only a few soils. In the present studies, therefore, 35 soils from a number of field locations of the Punjab province, were included to test this approach. Evaluation of the index has been made by comparing it with the performance of incubation test and various conventional chemical methods in relation to yield and N uptake by wheat plants.

Materials and methods

Soils

Surface soil samples (0–15 cm) used in this study were air dried soon after collection from the field. After passing through a 2-mm sieve, the soils were then analysed for some of their

physio-chemical properties. The pH of the soils ranged from 7.4–7.9, total N 0.033–0.099 per cent, organic matter 0.49–1.57 per cent and clay 12.0–37.4 per cent. pH of the soil paste was measured by glass electrode, organic matter was determined by dichromate oxidation¹³ and total N by salicylic acid method².

Soil extraction procedure

i) *Extraction with 2 N KCl*. Five-g soil samples (in triplicate) were shaken with 2 N KCl for 1 h and the soil extracts were then analysed for $\text{NO}_3\text{-N}$ by steam distillation with Devarda's alloy and MgO^3 .

ii) *Extraction with different concentrations of KMnO_4 in 1 N H_2SO_4 following pre-extraction with 1 N H_2SO_4* . The extraction procedure was almost the same as proposed by Stanford and Smith¹² but the extracts recovered by centrifugation after the initial shaking of the soil samples with 1 N H_2SO_4 , were also retained for their $\text{NH}_4\text{-N}$ analysis. Acid permanganate extracting solutions were prepared on the day of extraction by dissolving appropriate amounts of KMnO_4 in 1 N H_2SO_4 to get the final KMnO_4 normality of 0.05, 0.1 and 0.2. The extraction procedure was as follows:

Two-g soil samples (in triplicate) taken in centrifuge tubes were shaken with 50 ml of 1 N H_2SO_4 for 1 h at room temperature. After centrifugation the supernatants were analysed for $\text{NH}_4\text{-N}$ by steam distillation using NaOH as the alkalizer². The soil residues were then shaken with 50 ml of acid KMnO_4 solution for 1 h at room temperature and then centrifuged. The supernatants collected were analysed for $\text{NH}_4\text{-N}$ as described before.

Nitrate-N extracted by 2 N KCl and $\text{NH}_4\text{-N}$ extracted by 1 N H_2SO_4 will be collectively designated as "mineral N" unless otherwise mentioned. Ammonium-N released owing to the oxidation of soil organic matter by acid permanganate solution will be referred as "mineralizable N" and the concentrations of KMnO_4 in the extractant will be specified.

Incubation method

Portions of 50 g of soils were taken in flat bottomed plastic vessels with lids having a central hole to provide gaseous exchange. Soil samples in triplicate were brought to their respective 75 per cent field capacities by the addition of distilled water and incubated at $30 \pm 1^\circ\text{C}$ for 4 weeks. Soil moisture was maintained at 75 per cent field capacity by weighing the bottles and adding distilled water subsequently, at frequent intervals during the course of the experiment. At the end of the incubation period, the soils in the plastic vessels were thoroughly mixed and subsamples were extracted with 2 N KCl. Mineral N of incubation test was determined by steam distillation of the soil extract with Davarda's alloy and MgO^3 .

Pot culture

Triplicate 1.5 kg subsamples of each soil were filled in polythene lined plastic pots and then brought to their respective field capacities by the addition of distilled water. Nine seeds of wheat (*Triticum aestivum* L.; cv. Sandal) were sown in each pot and the stand thinned to 5 seedlings thereafter. Soil moisture was maintained at field capacities of the soils by daily addition of distilled water throughout the plant growth period. The plants were harvested 40 days after germination and rinsed thoroughly in distilled water. Plant tops were oven dried at 70°C for 48 h and total N in the ground plant material was determined by Kjeldahl method.

Results and discussion

A comparison of different N availability indexes as estimated by biological and chemical methods and their relation to dry matter yield and N uptake by wheat plants is given in Table 1.

Table 1. Correlation coefficients of different N availability indexes vs. dry matter yield and N uptake by wheat (n = 35)

Measure of available N	Correlation coefficient (r)	
	Dry matter	N uptake
1. Soil incubation	0.44**	0.82**
2. Mineral N (by 2 N KCl and 1 N H ₂ SO ₄), plus mineralizable N extracted by acidic:		
i) 0.05 N KMnO ₄	0.53**	0.72**
ii) 0.1 N KMnO ₄	0.49**	0.68**
iii) 0.2 N KMnO ₄	0.52**	0.66**
3. Mineral N (2 N KCl)	0.13 ^{ns}	0.62**
4. Total soil N	0.38*	0.38*
5. Organic matter	0.46**	0.38*

*Significant at P 0.05, **Significant at P 0.01, ns Non-significant

Incubation method

Mineral N after 4 weeks of soil incubation ranged from 18 to 77 mg/kg of soil. The correlation between mineral N of incubation test and N uptake by wheat plants has been shown in Fig. 1 which is quite high ($r = 0.82$, $P < 0.01$). This high association clearly indicates soil incubation method to be a reliable index of soil N availability. This has already been confirmed¹.

Chemical methods

Mineralizable N released by 0.05 N KMnO₄ in 1 N H₂SO₄ did not correlate well with the yield and N uptake by the plants. Mineralizable N extracted with higher concentrations of acid KMnO₄ solution correlated significantly ($P < 0.05$ in most of the cases) with both the plant yield and N uptake but the correlation in either case was relatively poor. From these results it appears that the determination of soil mineralizable N by acid permanganate extraction does not provide a good index of available N. The results of Castellanos and Pratt⁴ also demonstrated that the amounts of N released by KMnO₄ were not significantly correlated with available N of the manures estimated as their original mineral N plus net N mineralized during a 10-month period of green house cropping. In the present studies when the values of mineral N were included with those of mineralizable N (released by different concentrations of acid KMnO₄ solution), the correlation with the plant yield and N uptake values was much improved (Table 1). The improved relationship could be explained on the basis that the initial mineral N content of the soils contributes in meeting the N requirements of plants.

Mineralizable N has been reported to be good index of soil N availability¹. However, this index is open to many criticisms. One of them is

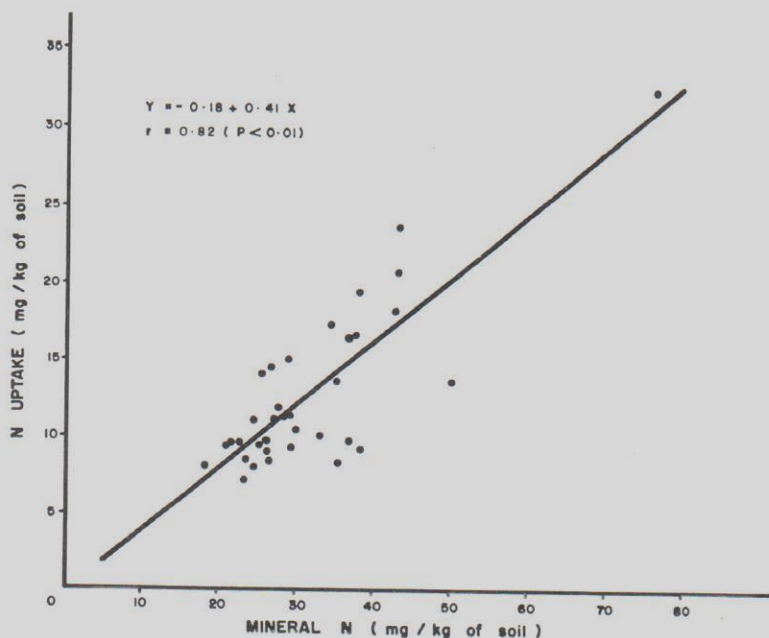


Fig. 1. Relationship between N uptake by wheat and mineral N produced by aerobic incubation of soils at $30 \pm 1^\circ\text{C}$ for 4 weeks.

that mineralizable N does not provide an estimate of the amount of mineral N present in the sample before incubation and takes no account of mineral N which has accumulated during fallow or from fertilizer applications¹. The results of the present study also emphasise that the initial mineral N (2 N KC1) content of soils are also worthy of consideration while devising some method for obtaining N availability index. Certain other investigators have interpreted similar findings indicating that initial mineral N should not be overlooked¹¹. They suggested that if an appreciable amount is present in the root zone at the beginning of a cropping season, its potential contribution, as well as that of the organic soil N must be considered in making a valid prediction of N fertilizer needs. For crops like winter wheat, potatoes and sugar beet, soil analysis for mineral N has been experimentally demonstrated to be a means of better assessing optimum fertilizer N requirements⁸. Sahrawat and Burford⁹ also pointed out that nitrates are not included in the available N pool when the alkaline permanganate method is used for determination of available soil N. According to them this may not be of importance for wetland soil where nitrate is unlikely to be present. But nitrate may accumulate in appreciable amounts in upland soils, and it may contribute significantly to a soil pool of available N.

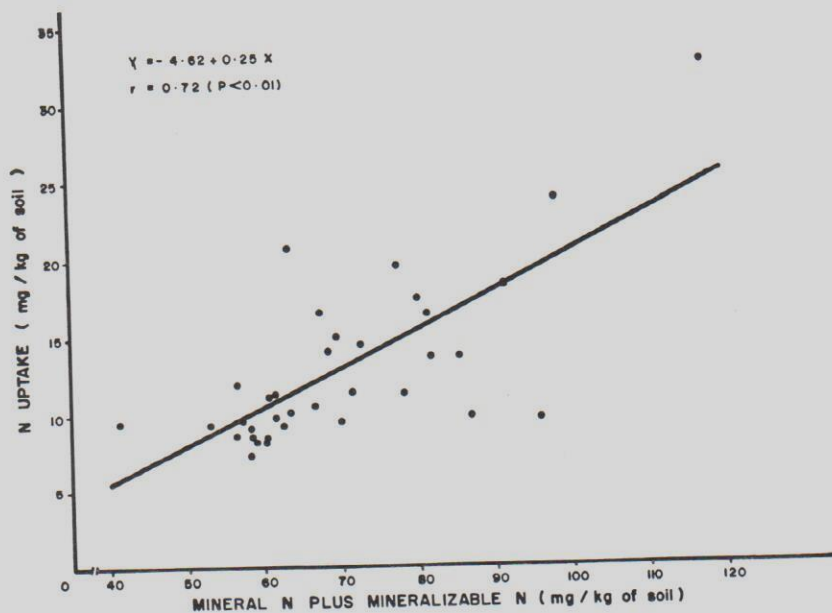


Fig. 2. Relationship between N uptake by wheat and soil available N values estimated as mineral N, plus mineralizable N by 0.05 N KMnO₄ in 1 N H₂SO₄.

The present data also indicated that in case of the method involving estimation of mineral N plus mineralizable N, 0.05 N KMnO₄ concentration in 1 N H₂SO₄ was quite suitable for extracting mineralizable N of soils ($r = 0.72$, $P < 0.01$, Fig. 2). Using higher concentrations of KMnO₄ in the extraction solution could not further improve the correlation values (Table 1). Stanford and Smith¹² while presenting a new chemical index of soil N availability based on acid permanganate oxidation, proposed that either of the two concentrations of KMnO₄ in 1 N H₂SO₄ *i.e.* 0.05 N or 0.1 N could be used for extracting the soils.

The correlation coefficients of mineral N (2 N KCl), total soil N and organic matter *vs.* dry matter yield and N uptake by plants were found to be either nonsignificant or quite poor. The present studies revealed that after soil incubation the method involving the estimation of mineral N plus mineralizable N proved to be a useful index of available N. Since the chemical method proposed is less time consuming and when compared with other chemical methods gives good correlation with N uptake by wheat, it deserves consideration to be used as a routine test for assessing N availability in soils.

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